

High and Low Level Mupirocin Resistance among Clinical Isolates of Methicillin Resistant *Staphylococcus aureus* (MRSA)

Zainab Yousaf¹, Farhan Rasheed², Muhammad Touqeer Hanif³

¹Department of Pathology Farooq Hospital, Westwood Colony, Lahore, ² Department of Pathology Allama Iqbal Medical College Lahore, ³ College of Allied Health Sciences, Akhtar Saeed Medical & Dental College

ABSTRACT

Objective: *Staphylococcus aureus* is a significant source of nosocomial infections, causing illnesses ranging from minor to life-threatening septic shock. Routine culturing and specific testing can confirm the presence of the bacteria. Antibiotic resistance is a major concern, and methicillin-resistant *Staphylococcus aureus* (MRSA) is particularly dangerous as it has mutations in the *mecA* gene, making it resistant to beta-lactam antibiotics. Mupirocin is an effective antibiotic against MRSA, binding to isoleucyl transfer-RNA synthetase to inhibit protein and RNA synthesis. In this study, two different doses (5 µg and 200 µg) of mupirocin were tested against MRSA.

Methodology: This study aimed to determine the antimicrobial susceptibility of methicillin-resistant *Staphylococcus aureus* (MRSA) against two different doses of mupirocin. Clinical samples were collected in the duration of one year from different wards of Jinnah Hospital Lahore and processed in microbiology laboratory of Allama Iqbal Medical College and after confirmation of *Staphylococcus aureus*, the modified Kirby Bauer method was used to test antimicrobial susceptibility. MRSA was confirmed by resistance to ceftiofuran. High and low-level doses of mupirocin were applied, and zone sizes were measured.

Results: Total 172 isolated methicillin resistant *Staphylococcus aureus* (MRSA) from all age groups and genders were tested against two doses of mupirocin. 97.10% MRSA were sensitive against both doses (5 µg and 200 µg) of mupirocin.

Conclusion: After the resistance of all beta lactam drugs, mupirocin can be very helpful topically in the treatment of MRSA infections.

Key Words: MRSA, Mupirocin

Authors' Contribution:

^{1,2}Conception; Literature research; manuscript design and drafting; ^{2,3} Critical analysis and manuscript review; ³Data analysis; Manuscript Editing.

Correspondence:

Name: Zainab Yousaf
email: zainabyousaf00@gmail.com

Article info:

Received: April 18, 2023
Accepted: December 20, 2023

Cite this article. Yousaf Z, Rasheed F, Hanif TM. High and Low-Level Mupirocin Resistance among Clinical Isolates of Methicillin Resistant *Staphylococcus Aureus* (MRSA). J Islamabad Med Dental Coll. 2023; 12(4): 250-254
DOI: <https://doi.org/10.35787/jimdc.v12i4.983>

Funding Source: Nil
Conflict of interest: Nil

Introduction

Staphylococcus aureus is a significant threat to humans, as they are a major source of nosocomial infections. Nasal carriers are a significant source of infection spread in hospitals, particularly among immunocompromised patients. These Gram-

positive cocci also cause many illnesses like folliculitis to life-threatening septic shock. One great thing is that laboratories can detect it quickly and minimize the complications due to these bugs.^{1,2} Another essential addition to its threatening ability is its resistance for many common antibiotics.⁴

Antibiotic resistance is a threat causing alarm for humankind, but *S. aureus* becomes more dangerous than others when it gains drug resistance. *S. aureus*, becomes more dangerous when it gets mutation in specific gene *mecA* and survives in the presence of all beta-lactam drugs. Due to this new adaptation, the treatment of *S. aureus* infections is complicated. This mutated bug is known as Methicillin-Resistant *Staphylococcus aureus* is known as “MRSA”. Many methods for its confirmation, like, detection of PBP2 proteins, the Use of selective chrome agar and genes amplification through PCR, are available but, the most economical method is the cefoxitin disc diffusion method.^{3,5}

Few drugs act as the last hope to kill this highly resistant infectious agent. Among these, mupirocin is available with two different dose levels to cure infections. Mupirocin is an effective agent against MRSA. This antibiotic acts by binding with the isoleucyl transfer-RNA (tRNA) synthetase. Then inhibition of the action of these enzymes causes failure in the conversion and isoleucine and tRNA, hence no protein and RNA synthesis.^{4,6}

Mupirocin is a naturally synthesized antibiotic and is active against many gram-positive and a few gram-negative bacteria.⁷ It has a unique structure and is obtained with fermentation by *Pseudomonas fluorescens*. It is already available in the market for tropical use and provides a potential role in eradicating the nasal carriage when administered intranasal. The high metabolism of this drug is a significant hurdle in its use against systemic infections.⁸

Methodology

The ethical review board of Allama Iqbal Medical College and Jinnah Hospital Lahore given the approval of this study in 39th meeting. MRSA from all clinical samples, age groups, and genders were included in the study, and methicillin-sensitive *Staphylococcus aureus* (MSSA) were excluded. Samples were collected from different wards and

cultured on blood and MacConkey agar. After 48 hours of incubation, colonial morphology was noted, and gram staining was performed. Catalase test was done to differentiate between *Staphylococcus* and *Streptococcus*. Coagulase, DNase, and mannitol salt agar tests were performed to confirm *Staphylococcus aureus*.

After confirming *Staphylococcus aureus*, antimicrobial susceptibility testing was performed with the modified Kirby Bauer method. Isolates resistant to cefoxitin were labelled as methicillin-resistant *Staphylococcus aureus* (MRSA). After the confirmation of MRSA, high (MUP-200 µg) and low-level (MUP-5 µg) doses of mupirocin were applied to analyse the antimicrobial susceptibility against MRSA using the modified Kirby Bauer method. Plates were incubated for 24 hours at 37°C, and zone sizes were measured. Zone size for the sensitive isolates of MRSA for 5µg mupirocin disc was 14 mm, and for the 200µg disc was 30 mm. 5µg is considered low-level dose and 200µg were considered high dose mupirocin. All data were recorded and analysed by SPSS.

Results

Among 172 samples 39.5% and 60.5% were from female and male respectively as shown in table 1: All patients were divided into four age groups, minimum age was 1-year-old and maximum was 80 years old. Table 2 is the representation of population in all age groups and among them it was highest in between 21-40 years.

Gender	Frequency	Percent
Female	68	39.5
Male	104	60.5
Total	172	100.0

Frequency of patients in different age groups		
Age Groups	Frequency	Percent
1-20 years	28	16.3
21-40 years	82	47.7
41-60 years	48	27.9
61-80 years	14	8.1
Total	172	100.0

Frequency of MRSA isolated from different clinical specimens		
Specimens	Frequency	Percent
Blood	3	1.7%
CVP Tip	3	1.7%
Fluid	6	3.5%
HVS	2	1.2%
Nasal Swab	2	1.2%
Pus	96	55.8%
Sputum	7	4.1%
Tissue	1	.6%
Tracheal Secretions	5	2.9%
Urine	2	1.2%
Wound Swab	45	26.2%
Total	172	100.0%

Antimicrobial susceptibility testing was performed on all isolates and results were noted for all antibiotics low and high dose of mupirocin. Table 4 is the representation of all antibiotics except both doses of mupirocin. MRSA were isolated from different clinical specimens and table 3 is the representation of it. MRSA were highest in pus and wound swabs, 55.8% and 26.2% respectively. As two doses of mupirocin were applied, low dose (MUP-5) and high dose (MUP-200) on all the isolated MRSA.

Drug	Sensitive	Resistant
Amoxicillin/Clavulanic Acid	0%	100%
Methicillin	0%	100%
Ciprofloxacin	6.4%	93.6%
Gentamicin	37.8%	62.2%
Amikacin	45.3%	54.7%
Erythromycin	24.4%	75.6%
Clindamycin	45.9%	54.1%
Erythromycin	24.4%	75.6%
Fusidic Acid	59.3%	40.7%
Co-trimoxazole	12.8%	87.2%
Doxycycline	48.8%	51.2%
Chloramphenicol	54.1%	45.9%
Vancomycin	99.4%	0.6%
Linezolid	100%	0%

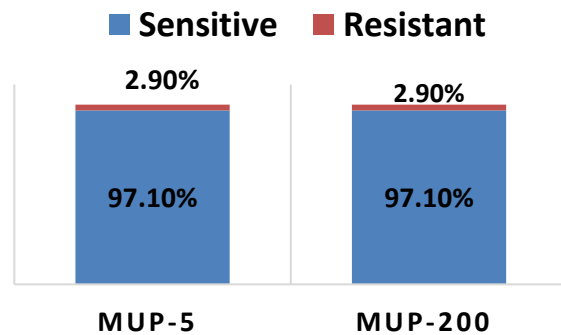


Fig. 1: Susceptibility of mupirocin (low and high dose)

Discussion

In our study, both the 5 µg and 200 µg doses of mupirocin exhibited high efficacy, demonstrating sensitivity in approximately 97.10% of MRSA isolates. It's a positive finding, suggesting the effectiveness of mupirocin in our specific context. However, when we consider the broader spectrum illuminated by other studies, nuances emerge. The global overview⁹ indicates a variable prevalence of mupirocin resistance worldwide, ranging from 7.6% to 13.8%. This suggests that while mupirocin may work well in some regions, resistance is a

noteworthy concern on a global scale. The longitudinal study¹⁰ spanning four years raises concerns about the persistent rise in mupirocin resistance despite ongoing infection control efforts. It prompts us to reflect on the challenges of maintaining the efficacy of this antimicrobial agent over time.

The genetic insights provided in a study¹¹ underscore the complexity of mupirocin resistance mechanisms. The study reveals a global emergence of resistance genes, emphasizing the adaptive nature of bacteria in response to antimicrobial agents. On a more localized scale, the study involving medical students¹² indicates a low risk of MRSA and mupirocin resistance within this demographic. However, the study in Malaysia¹³ highlights higher resistance rates in specific MRSA types, signalling the importance of tailoring interventions based on regional dynamics.

In the hospital setting¹⁴, the observed 13% overall mupirocin resistance rate necessitates regional vigilance. Meanwhile, the evolving molecular epidemiology in Singapore¹⁵ underscores the need for continuous surveillance to adapt to changing MRSA trends over time. In summary, while our study reflects positive outcomes for mupirocin in our specific setting, the global and temporal perspectives provided by other studies emphasize the need for a nuanced and adaptive approach. Mupirocin resistance is not only a regional concern but also a dynamic global challenge that warrants ongoing attention and strategic antimicrobial stewardship.

Conclusion

High susceptibility pattern of mupirocin in MRSA showed that, it can be used to cure infections due to methicillin resistant *Staphylococcus aureus* (MRSA) especially in soft tissues and skin wounds. This study was limited to susceptibility testing through the disc diffusion method.

References

1. Klevens RM, Edwards JR, Tenover FC, McDonald LC, Horan T, Gaynes R. Changes in the Epidemiology of Methicillin-Resistant *Staphylococcus aureus* in Intensive Care Units in US Hospitals, 1992-2003. *Clinical Infectious Diseases*. 2006 Feb 1;42(3):389-91. <https://doi.org/10.1086/499367>
2. Patino S, Alamo L, Cimino M, Y. Casart, Bartoli F, Garcia MJ, et al. Autofluorescence of Mycobacteria as a Tool for Detection of Mycobacterium tuberculosis. *Journal of Clinical Microbiology*. 2008 Oct 1;46(10):3296-302 <https://doi.org/10.1128/jcm.02183-07>
3. Chemidlin Prévost-Bouré N, Christen R, Dequiedt S, Mougél C, Lelièvre M, Jolivet C, et al. Validation and Application of a PCR Primer Set to Quantify Fungal Communities in the Soil Environment by Real-Time Quantitative PCR. Yu JH, editor. *PLoS ONE*. 2011 Sep 8;6(9):e24166. Moellering Jr, R. C. (2010). MRSA: the first half century. *Journal of antimicrobial chemotherapy*, 65(suppl_3), iii3-iii14. <https://doi.org/10.1093/jac/dkq303>
4. Apple FS, Jesse RL, Newby LK, Wu AHB, Christenson RH. National Academy of Clinical Biochemistry and IFCC Committee for Standardization of Markers of Cardiac Damage Laboratory Medicine Practice Guidelines: Analytical Issues for Biochemical Markers of Acute Coronary Syndromes. *Circulation*. 2007 Apr 3;115(13). <https://doi.org/10.1373/clinchem.2006.084715>
5. Bamidele O, Jiang ZD, Dupont H. Occurrence of putative virulence-related genes, *aatA*, *aggR* and *aaiC*, of Enteroaggregative *Escherichia coli* (EAEC) among adults with travelers' diarrhea acquired in Guatemala and Mexico. *Microbial Pathogenesis*. 2019 Mar;128:97-9. <https://doi.org/10.1016/j.micpath.2018.12.030>
6. Silva M, Milanese G, Seletti V, Ariani A, Sverzellati N. Pulmonary quantitative CT imaging in focal and diffuse disease: current research and clinical applications. *The British Journal of Radiology*. 2018 Jan 12;20170644. <https://doi.org/10.1259%2Fbjr.20170644>
7. Stutz A, Kolbe CC, Stahl R, Horvath GL, Franklin BS, van Ray O, et al. NLRP3 inflammasome assembly is regulated by phosphorylation of the pyrin domain. *The Journal of Experimental Medicine* [Internet]. 2017 Jun 5 [cited 2022 May 17];214(6):1725-36. <https://pubmed.ncbi.nlm.nih.gov/28465465/>
8. Dadashi M, Hajikhani B, Darban-Sarokhalil D, van Belkum A, Goudarzi M. Mupirocin Resistance in *Staphylococcus aureus*: A Systematic Review and

- Meta-Analysis. *Journal of Global Antimicrobial Resistance*. 2019 Aug;20.
<https://doi.org/10.1016/j.jgar.2019.07.032>
9. K. Szymanek-Majchrzak, Kosiński JW, Krzysztof Żak, Sulek K, Andrzej Młynarczyk, Grażyna Młynarczyk. Prevalence of methicillin resistant and mupirocin-resistant *Staphylococcus aureus* strains among medical students of Medical University in Warsaw. 2019 Jan 1;39–48.
<https://doi.org/10.32394/pe.73.05>
 10. Eltringham I. Mupirocin resistance and methicillin-resistant *Staphylococcus aureus* (MRSA). *Journal of Hospital Infection*. 1997 Jan;35(1):1–8.
[https://doi.org/10.1016/s0195-6701\(97\)90162-6](https://doi.org/10.1016/s0195-6701(97)90162-6)
 11. Ghasemzadeh-Moghaddam H, van Belkum A, Hamat RA, van Wamel W, Neela V. Methicillin-Susceptible and -Resistant *Staphylococcus aureus* with High-Level Antiseptic and Low-Level Mupirocin Resistance in Malaysia. *Microbial Drug Resistance*. 2014 Oct;20(5):472–7.
 12. Chen S, Jin Y, Lin C, Hao Z, Duan J, Guo Y, et al. Low prevalence of mupirocin resistance among *Staphylococcus aureus* clinical isolates from a Chinese tertiary hospital. *Journal of Medical Microbiology*. 2019 Feb 1;68(2):201–5.
<https://doi.org/10.1099/jmm.0.000911>
 13. Hon PY, Koh TH, Tan TY, Krishnan P, Leong JW, Jureen R, Chan J, Tee NW, Muruges J, Chan KS, Hsu LY. Changing molecular epidemiology and high rates of mupirocin resistance among methicillin-resistant *Staphylococcus aureus* in Singaporean hospitals. *Journal of Global Antimicrobial Resistance*. 2014 Mar 1;2(1):53-5
<https://doi.org/10.1016/j.jgar.2013.10.002>